# PREPARATION OF DEUTERIUM LABELLED α-AMINO-3-HYDROXY5-METHYLISOXAZOLE-4-PROPIONIC ACID (AMPA)

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#### SUMMARY

The preparation of  $\alpha$ -amino-3-hydroxy-5-methylisoxazole-4-propionic acid (AMPA) with deuterium  $\alpha$  to the carboxylic acid group (2) and in the exocyclic methyl group (5), respectively, are described. The deuterium labelled AMPA (2) was synthesized by hydrolysis and decarboxylation of the corresponding di-ester (1), whereas the deuterium labelled AMPA (5) was prepared via a catalytic deuteration of the bromoderivative (3).

Key Words: AMPA, Glutamic Acid Agonist, Deuterium

# INTRODUCTION

L-Glutamic acid (GLU) is a putative excitatory transmitter in the mammalian central nervous system based on *in vivo* experiments. 1-4 The understanding of the mechanism involved in receptor mediated excitations produced by GLU, will be facilitated by the identification and characterization of the physiological GLU-receptor using ligand binding techniques. Several attempts have been made to identify the GLU-receptor using [<sup>3</sup>H]GLU and [<sup>3</sup>H]kainic acid (KAIN), a structural analogue of GLU (Fig. 1), as ligands. 5-18 However, the characteristics of the binding sites so

Figure 1. The structures of GLU, AMPA, and KAIN

far detected are inconsistent with the characteristics of a physiological GLU receptor.

The title compound  $\alpha$ -amino-3-hydroxy-5-methylisoxazole-4-propionic acid (AMPA, Fig. 1) has proved to be a very potent and selective GLU-agonist in  $in\ vivo$  experiments. However, when tested as an inhibitor of [ $^3$ H]GLU and [ $^3$ H]KAIN binding AMPA was virtually inactive. Consequently, tritium labelled AMPA is an important tool in the search for the physiological GLU-receptor. The present paper describes the synthesis of deuterium labelled AMPA, which can also be applied for the synthesis of tritiated AMPA.

#### RESULTS AND DISCUSSION

AMPA was deuterium labelled i) in the position  $\alpha$  to the carboxylic acid group and ii) in the exocyclic methyl group.

The syntheses are outlined in scheme 1.

Treatment of 1 with 6 M  $^2$ HCl in  $^2$ H $_2$ O gave, after removal of labile deuterium with water, labelled AMPA hydrochloride (2) with deuterium in the  $\alpha$ -position to the carboxylic acid group.

Treatment of 1 with N-bromosuccinimide (NBS) under free radical conditions was shown by  $^{1}{\rm H}$  NMR spectroscopy to give 3 as the major product in contrast to the findings that NBS bromination of 4,5-dimethyl-3-methoxyisoxazole resulted in bromination in the 4-me-

## Scheme 1.

thyl group. <sup>21</sup> Catalytic low pressure deuteration of 3 using triethylamine (TEA) as a base afforded the deuterated compound 4. Deprotection of 4 using 48 % aqueous HBr gave labelled AMPA (5) with deuterium in the exocyclic methyl group.

In order to investigate the stability of deuterium in 2 the compound was dissolved in water and kept at ambient temperature for 36 h. The  $^1\mathrm{H}$  NMR spectrum of the evaporated solution showed a minor content of non-labelled AMPA, which was not quantified. The instability of 2 was confirmed by experiments with the tritium labelled compound.  $^{20}$  In these experiments 27 per cent of tritium were labilized on storage of the tritium labelled compound in aqueous solution for 2 days at 3  $^{\mathrm{O}}\mathrm{C}$ .

The deuterium in compound 5 was considered to be permanent, as the deuterium in compound 4 was stable in refluxing 48 % aqueous  ${\tt HBr.}$ 

## EXPERIMENTAL

Melting points, determined in capillary tubes, are corrected. Analyses indicated by elemental symbols were within  $\pm 0.4$  % of the theoretical values and were performed by Mr. P. Hansen, Chemical Laboratory II, University of Copenhagen.

A JEOL JMN-C-60HL (60 MHz)  $^1$ H NMR instrument was used.  $^1$ H NMR spectra were recorded using TMS as an internal standard, except for the compounds dissolved in  $\rm D_2O$  where DSS was used. TLC and column chromatography were accomplished by using silica gel  $\rm F_{254}$  plates (Merck) and silica gel, 0.063-0.100 (Woelm), respectively.

 $[\alpha^{-2}H]\alpha$ -Amino-3-hydroxy-5-methylisoxazole-4-propionic acid, hydrochloride (2). A solution of 1  $^{21}$  (170 mg; 0.05 mmol) in 6 M  $^{2}$ HCl in  $^{2}H_{2}O$  (10 ml) was refluxed for 2 h. Evaporation followed by evaporation twice from  $H_{2}O$  and recrystallization (glacial acetic acid:water) gave 2 (80 mg; 72 %). TLC [butanol-glacial acetic acid-water (4:1:1)]: comparison with an authentic sample of AMPA  $^{21}$  corresponded.  $^{1}H$  NMR ( $D_{2}O$ ):  $\delta$  4.75 ( $c\alpha$ . 5 H, s), 3.00 (2 H, s), 2.30 (3 H, s).

Ethyl  $\alpha$ -ethoxycarbonyl- $\alpha$ -acetylamino-3-methoxy-5-bromomethylis-oxazole-4-propionate (3). To a solution of 1  $^{21}$  (2.05 g; 6 mmol) in CCl<sub>4</sub> (12 ml) was added NBS (1.07 g; 6 mmol) and benzoyl peroxide (10 mg). The reaction mixture was refluxed for 30 min. Filtration and evaporation followed by column chromatography [methylene chloride - 2-butanone (95:5)] and recrystallization (ethylacetate-light petroleum) gave 3 (0.8 g; 32 %). M.p. 147.0-147.5  $^{\circ}$ C. Anal. C<sub>15</sub>H<sub>21</sub>BrN<sub>2</sub>O<sub>7</sub>: C, H, Br, N. IR (KBr): 3360 (m), 2980 (m), 1730 (s), 1660 (s), 1530 (s), 1500 (m) cm<sup>-1</sup>.  $^{1}$ H NMR (CDCl<sub>3</sub>):  $^{\circ}$ 6.7 (1 H, broad s), 4.5-4.0 (6 H, m containing a s), 3.95 (3 H, s), 3.45 (2 H, s), 2.05 (3 H, s), 1.30 (6 H, t, J 7 Hz).

Ethyl  $\alpha$ -ethoxycarbonyl- $\alpha$ -acetylamino-3-methoxy-5-[ $^2$ H]methylis-oxazole-4-propionate (4). To a solution of 3 (500 mg; 1.2 mmol)

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α-Amino-3-hydroxy-[ $^2$ H]methylisoxazole-4-propionic acid (5). A solution of 4 (130 mg; 0.4 mmol) in 48 % aqueous HBr (13 ml) was refluxed at 140  $^{\circ}$ C for 15 min. After evaporation and recrystallization (glacial acetic acid:ether) 5 (90 mg; 88 %) was obtained as s hydrobromide. TLC [butanol-glacial acetic acid-water (4:1:1)]: comparison with an authentic sample of AMPA  $^{21}$  corresponded.  $^{1}$ H NMR (D<sub>2</sub>O):  $\delta$  4.75 ( $\epsilon$ a. 5 H, s), 4.30 (1 H, t,  $\epsilon$  6.0 Hz), 3.00 (2 H, d,  $\epsilon$  6.0 Hz), 2.4-2.2 (2 H, m).

- Curtis, D.R. and Johnston, G.A.R. Ergeb. Physiol. Biol. Chem. Exp. Pharmakol. 69:97 (1974).
- 2. Krnjević, K. Physiol. Rev. 418 (1974).
- Curtis, D.R. In Filer, L.J., Jr., Garattini, S., Kare, M.R., Reynolds, W.A. and Wurtman, R.J., Eds., Glutamic Acid: Advances in Biochemistry and Physiology, Raven Press, New York 1979, p. 163.
- Nistri, A. and Constanti, A. Prog. Neurobiol. (Oxford)
  13:117 (1979).
- 5. Michaelis, E.K., Michaelis, M.L. and Boyarsky, L.L. Biochim. Biophys. Acta 367:338 (1974).

- 6. Michaelis, E.K. Biochem. Biophys. Res. Commun. 65:1004 (1975).
- 7. Roberts, P.J. Nature (London) 252:399 (1974).
- 8. De Roberto, E. and Fiszer de Plazas, S. J. Neurochem. 26:1237 (1976).
- 9. Foster, A.C. and Roberts, P.J. J. Neurochem. 31:1467 (1978).
- 10. Baudry, M. and Lynch, G. Eur. J. Pharmacol. 57:283 (1979).
- 11. De Barry, J., Vincendon, G. and Gombos, G. FEBS Lett. 109:175 (1980).
- 12. Biziere, K., Thompson, H. and Coyle, J.T. Brain Res. 183: 421 (1980).
- 13. Simon, J.R., Contrera, J.F. and Kuhar, M.J. J. Neurochem. 26:141 (1976).
- 14. London, E.D. and Coyle, J.T. Mol. Pharmacol. 15:492 (1979).
- 15. Vincent, S.R. and McGeer, E.G. Life Sci. 24:265 (1979).
- 16. Schwarz, R. and Fuxe, K. Life Sci. 24:1471 (1979).
- 17. Henke, H, Neurosci. Lett. 14:247 (1979).
- 18. Honoré, T., Lauridsen, J. and Krogsgaard-Larsen, P. J. Neurochem. 1980 (in press).
- 19. Krogsgaard-Larsen, P., Honoré, T., Hansen, J.J., Curtis, D.R. and Lodge, D. Nature (London) 284:64 (1980).
- 20. The Radiochemical Centre, Amersham, England. Personal Communication.
- 21. Honoré, T. and Lauridsen, J. Acta Chem. Scand. <u>B34</u>:235 (1980).